

الوصف النباتي والتركيب الكيميائي والتأثيرات العلاجية لنبات المرسين حنان محمد ميلاد أبوزويته ، وفوزيّة عامر أبوزيد ، وآمنة المهدي لحضريري ، وآمنة صالح القذافي ، وإيمان ضو احمد .

المـاـخـصـ:

ستسلط هذه المراجعة على المكونات الكيميائية ، وهي وصف تفصيلي للشكل والتأثيرات الدوائية والعلاجية لنبات *Myrtus L.* *communis*. هذا النبات هو عشب دائم الخضرة من العائلة الكافورية Myrtaceae ، ينتشر على نطاق واسع في جميع أنحاء العالم. حيث كانت نسبة الكربوهيدرات (88,69%) والبروتين الخام (5,97%) والزيت الخام (3,59%) ومحتوى الرطوبة (63,67%) وإجمالي الرماد (1,75%) بالإضافة إلى أنه يحتوى على عناصر معدنية ومحتويات نشطة بيولوجيا . وله العديد من الأنشطة الدوائية مثل : مضادات الجراثيم ، مضادات الفطريات ، مضادات السرطان ، مضادات الفيروسات ، مضادات الأكسدة ومضادات السكر والحماية الكبدية والعصبية ، وما إلى ذلك من مستخلصات نباتية مختلفة.

*Myrtus communis L.*phytochemistry, ethnobotany and pharmacology: A review

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Abstract

This review will highlight the chemical constituents and detail description of morphology and the pharmacological and therapeutic effects of *Myrtus communis*. This plant commonly known as myrtle, is an evergreen and aromatic herb of the family Myrtaceae, it is widely distributed throughout the world. *M. communis* was reported to contain carbohydrates (88.69%), crude protein (5.97%), crude oil (3.59%), moisture content (63.67%) and total ash (1.75%) and mineral elements and many other bioactive contents. Many

pharmacological activities viz., antibacterial, antifungal, anticancer, antiviral, antioxidant, antidiabetic, hepatoprotective and neuroprotective etc. have been reported in different plant extracts.

Keywords: *Myrtus communis* L., pharmacological activities, chemical constituents, ethnobotany, traditional uses.

Abbreviations

DPPH, 2,2-diphenyl-1-picrylhydrazyl; bw, body weight; ABTS, 2,2'-azino-bis(3-ethylbenzothiazoline-6-sulfonic acid; i.p., Intra-peritoneal injection; ER, Oesophageal Reflux; MBJ, Myrtle Berries Juice; HSV-1, Herpes Simplex Virus-1; CQ, Chloroquine, FRAP, Ferric Reducing Antioxidant Power Assay; MIC₅₀, Minimal Inhibitory Concentration.

Introduction

Myrtle (*Myrtus communis*) of the Myrtaceae family is an evergreen and aromatic shrub with numerous branches and stems [1]. Myrtaceae family contains 3,000 species naturally growing in subtropical and tropical regions. [2,3] It is native to North Africa, Southern Europe and West Asia. It is distributed in Southern America, Australia and North western Himalaya and widespread in the Mediterranean region [2].

For a long time, different parts of this plant such as branches, berries, leaves, seeds and fruits have been extensively used in traditional medicine in various doses form to treat haemorrhoids, diarrhoea, palpitations, conjunctivitis, peptic ulcers, leucorrhoea, pulmonary and skin disorders [4,5,6]. In addition to use as antibacterial, antifungal, anticancer, antiviral, antioxidant, antidiabetic, hepatoprotective and neuroprotective [5].

M. communis contained coumarins, myricetin, myrtucommulone A and B, myrtenol acetate, Myrtenol, limonene pinene, geraniol, p-cymene, phenylpropanoid, methyl eugenol phospholipids, phenolic compounds and essential oil [7,8].

Taxonomic Hierarchy

- Kingdom: Plantae
- Division: Spermatophytes
- Sub-Division: Angiosperms
- Class: Dicotyledone

- Order: Mytales
- Family: Myrtaceae
- Genus: *Myrtus*
- Species: *communis* [9]

Common Names

- South Arabia: Habb-ul-Aas, Hadass
- China: Xiang tao mu
- English: Myrtle
- Greek: Mirtia
- Italy: Mirto
- Persian: Barg-e-murad (leaves)
- Russia: Mirt
- Turkey: Mersin
- Urdu: Aass, Moorad
- Hindi: Sata Sova, Vilayati mehndi
- Sanskrit: Gandhamalati [10]
- Libya: Mersin [9]
- Iraq: Yas [11]

Botanical Description (Figures 1-5)

Leaves: are glossy, glabrous, dark green, 2.5-3.8 cm long, opposite, coriaceous, paired or whorled, ovate, lanceolate with stiff structure, aromatic.

Stem: 2.4-3 m high, its branches form a close full head, thickly covered with evergreen leaves.

Fruits: Berries are pea-sized, orbicular or ovoid ellipsoid, blue-black or white with hard kidney. The developed fruit is initially pale green, then turns deeper and finally becomes dark indigo when fully mature. They are bitter when unripe, sweet when ripe.

Seeds: Varying shapes and sizes 0.7-1.2 cm, the glabrous berry has rounded (vase-like) shaped with a swollen central part and remnants of persistent 4-5 partite calyx at the outer part [12-17].

Flowers: Star-like, white, with five sepals, five petals with glands and somewhat tomentose margin covered with fine hairs and a tufted mass of stamens [10].

Nutritional Value

Myrtus communis is a rich source of carbohydrates (88.69%), crude protein (5.97%), crude oil (3.59%), moisture content (63.67%) and total ash (1.75%) [18]. It contained mineral elements such as, calcium, sodium, potassium, magnesium, phosphorus, iron, zinc, aluminium, copper and plumbum [19].

Traditional uses

The leaves are useful in cerebral diseases especially epilepsy, stomach diseases [20,21], dyspepsia, liver diseases, rheumatism [21, 22], aphthae, eczema, pulmonary disorders [20,23], piles, sores [22], intertrigo, wounds, ulcers [24], stomatitis, deep inuses, uterine prolapse, leucorrhoea, internal ulceration, haemorrhage [25], inflammation, diarrhoea, hair fall, burns, herpes, palpitation, mennorrhagia [26], chronic bronchitis [27], abscess, sprain, diaphoresis [24] and chronic catarrah of bladder [27,28]. Various pharmacological actions of leaves are astringent, antiseptic [24], hypoglycaemic, laxative [27], analgesic [22,29], haemostatic [29], hair tonic [26] and stimulant [24].

Decoction of leaves and fruits is utilized orally for the cure of disbiosis, hypoglycaemia, stomachaches, cough constipation, poor appetite, externally for wound healing, urinary infections, enema and against respiratory diseases [30,3].

The berries are used in diarrhoea, dysentery, internal ulceration, rheumatism [22,21] foot ulcers, foetid ulcers, aphthae, deepsinuses, haemorrhages, leucorrhoea, lax vaginal walls [24], bronchitis [22], haemorrhoids [22], malaena, rhinitis, rectitis, conjuctivitis, piles, burns, dysurea, cough, epistaxis [29], earache, toothache, headache, palpitation [31], otorrhea [24], sprain, fractures, fever, polydipsia, burning micturition, scorpion sting, dandruff, melasma cholasma, menorrhagia, haemoptysis, uterine prolapse, rectal prolapse, eye ulcers, halitosis (bad breath), head, ulcers, vomiting, inflammations and gastric ulcer [32]. They are used as antiseptic, astringent [24], carminative [20,22,24], emmenagogue [22,24], demulcent, dessicant, analgesic, hair tonic, haemostatic [26], antiemetic, lithotripsic [33], cardiotonic, diuretic [22,29,33], anti-inflammatory [3], stomachic, brain tonic [22,33], haemostatic, nephroprotective, antidote [3], antidiaphoretic [29] and antidiabetic [27]. Different parts of the plant have been used in the food

industry, for example for flavouring meat and sauces, and in the cosmetic industry [27, 34].

In Turkish folk medicine, the fruits and leaves have been used in the treatment of urinary diseases, for healing wounds and as an antiseptic [35]. The oil of leaves and dried leaves of the plant are used to lower blood sugar in diabetic patients. In the Mediterranean regions, the tea is used to treat urinary tract infections and bladder [36]. In Libya, the plant is used for the treatment of cough, bronchitis, tuberculosis, lung complaints [37].

Chemical constituents

M. communis contains many active biological compounds which contribute significantly to the medicinal properties of the plant. Table 1 summarized the active constituents recorded from different parts of *M. communis*.

Table 1: Chemical constituents of *M. communis*

Constituents	Plant part	Reference
<ul style="list-style-type: none"> • Acylphloroglucinols: Myrtucommulone B,C,D,E and usnic acid derivative usnone A. 	The aerial parts	[38]
<ul style="list-style-type: none"> • Tannins, flavonoids such as quercetin, catechin and myricetin derivatives and volatile oils. • Alpha-pinene, 1V8-cineole, limonene and linalool. 	The leaves	[35,39] [40]
<ul style="list-style-type: none"> • The five terpenoid compounds (myrtenyl acetate, 1, 8-cineole, limonene, linalool). 	Leaf oil	[41]
Couenstittnts	Plant part	Reference
<ul style="list-style-type: none"> • Citric acid, malic acid, resin, tannin, fixed oil and sugar. • Flavonoids,anthocyanin arabinosides, anthocyanin glucosides. • Kaempferol, quercetin, myricetin 3-o-glucoside, 	Berries	[42,43] [44] [45]

myricetin 3, 3-di-o-galactoside, myricetin 3 rutinoside, aesculin, scopoletin, caffeic acid.		
<ul style="list-style-type: none"> Myricetin 3-o-rhamnoside or myricitrin, esculetin-6-o-glucoside or esculin, hesperetin 7-o-rhamnoglucoside or hesperidin, hesperetin-2-o-methylchalcone-4-o-rhamnoglucoside. 	Berries	[46]
<ul style="list-style-type: none"> The five terpenoid compounds (myrtenyl acetate, 1, 8-cineole, limonene, linalool). Flavonoids, such as alpha-pinene, 1V2-cineole, myrtenol, myrtenol, myrtenyl acetate, myrcene, Linalool and geraniol. 	Flowers	[41] [47]
<ul style="list-style-type: none"> Volatile oils, tannins, sugars, flavonoids and organic acids such as citric and malic acids. 14 fatty acids, oleic acid being the dominant fatty acid followed by palmitic acid and stearic acid. Flavonoids such as asqueretin, catechin, myricetin. 	Fruits	[35,33] [30] [48]
<ul style="list-style-type: none"> Tannins. Alkaloids, glycosides, reducing sugars Fixed oil. Gallic acids, phenolic acids, quercetin and patuletin. 	Roots	[31] [25] [21] [49]
Couenstittnts	Plant part	Reference
<ul style="list-style-type: none"> Ellagic acid and ellagitannins(eugeniflorin D₂ and oenothein B) Fatty oil (fixed oil) consisting of glycerides of oleic, linoleic, myristic, palmitic, linolenic and lauric acid 	Seeds	[50] [20,51]

Pharmacological activities

M. communis is reported for antimicrobial, antioxidant, Anticancer, Anti-inflammatory, Cardiovascular, Antidiabetic, Antinociceptive, Antidiarrheal Antiviral, Anthelmintic and other beneficial effects which are mentioned in Table 2.

Table 2: Pharmacological activities of *M. communis*

Activity	Dose	Extract/ Constituent	Model	Results	Reference
Antioxidant	DPPH: (4±0.3 to 21±0.1 g/ml), ABTS: 0.001 50±0.000 09 to 0.004 80±0.000 08 mg/ml	Methanol, chloroform, ethyl acetate and aqueous extracts	ABTS, DPPH, Hydroxyl radical scavenging activity, Metal chelating activity β - carotene/li noleic acid bleaching assay,, Reducing power,Ferr ic thiocyanat e test, <i>in vitro</i>	Ethyl acetate extract exhibited the highest activity in scavenging DPPH, ABTS, hydroxyl radical and reducing power	[52]
	In lipid peroxidation IC ₅₀ : 160 μ g/ml and 220 μ g/ml DPPH IC ₅₀ : 1.4 μ g/ml	Myricetin-3- ogalactoside and - rhamnoside, isolated from the leaves	Xanthine oxidase, lipid peroxidatio n and DPPH/ <i>in vitro</i>	Both compounds showed the most potent inhibitory effect in xanthine oxidase	[53]
Activity	Dose	Extract/ Constituent	Model	Results	Reference
Antioxidant	3 to 5 grams	Fruit extract	DPPH and β	Protect against	[54]

			-carotene-linoleic acid assays	lipid peroxidation and can scavenge free radicals	
	1 ml	Methanol of roots	DPPH, FRAP and Folin-Ciocalteu assays	This extract exhibited very good radical-scavenging activity.	[55]
Cardiovascular	0.04 to 12 mg/kg bw	Methanolic and ethyl acetate extracts of berries	In anaesthetized rats. Intravenous administration of both extracts	Decreased the maximum mean arterial blood pressure at 12 mg/kg, indicated that both extracts have lowering effect for blood pressure	[52]
	Relaxed phenylephrine (1µM) and K+ (80 mM)-induced contractions	Crude methanol extract	Isolated rabbit aorta preparations	Results were identical to verapamil, a standard calcium channel blocker	[56]
Anti-inflammatory	0.3-10 µg/ml	Ethanolic extract of leaves	6-keto-prostaglandin F1 α and [³ H]-arachidonic acid	Significant anti-inflammatory activity at the maximum	[57]

			metabolite production in keratinocytes	concentration	
Activity	Dose	Extract/ Constituent	Model	Results	Reference
Anti-inflammatory	1 and 2 ml/kg	Essential oil	Croton oil induced ear oedema and cotton pellet induced granuloma in mice	1. Significant ↓ ear oedema 2. Inhibit cotton pellet-induced granuloma and serum TNF- α and IL-6	[58]
Anti-inflammatory	0.5 to 4.5 mg/kg i.p.	Myrtucommulone (isolated from leaves)	Carrageenan-induced paw oedema and Pleurisy in mice	1. ↓ the growth of paw oedema in a dose-dependent manner 2. At 4.5 mg/kg i.p. myrtucommulone exerted potent anti-inflammatory effects in the pleurisy model	[59]
Anticancer	(1 and 3 μ g/ml)	Ethanolic extract	In vitro on HaCat keratinocytes by using the BrdU incorporation assay	The result showed that extract inhibit keratinocyte proliferation by 27% (1	[60]

				$\mu\text{g/ml}$ and 76% (3 $\mu\text{g/ml}$)	
		Essential oils	In two human cell lines HL- 60 and NB4	Is more effective on Ehrlich Ascites Carcinoma Cells in both <i>in vitro</i> and <i>in vivo</i> studies	[61]
Activity	Dose	Extract/ Constituent	Model	Results	Reference
Antibacterial	10,20, 40&80mg /ml	Hydroalcoholic of the leaves	Four pathogenic bacteria, <i>Staphylococcus</i> <i>aureus</i> , <i>Pseudomonas</i> <i>aeruginosa</i> , <i>Vibrio</i> <i>cholera</i> & <i>Escherichia coli</i>	The concentration of 80mg/ml showed the greatest effect on the <i>S. aureus</i> and <i>V. cholera</i>	[1]
Antibacterial	10 μl	Ethanolic and petroleum ether of fruits	<i>Enterobacter faecalis</i> , <i>Shigella Sonni</i> , <i>Micrococcus leutus</i> , <i>Bacillus subtilis</i> , <i>Staphylococcus aureus</i>	Ethanolic extract showed antibacterial activity against <i>E.</i> <i>faecalis</i> , <i>S.</i> <i>aureus</i> . Petroleum ether extract had no effect on tested bacteria.	[18]
	7.5 - 60 mg/ml	Essential oil of fruits	<i>Bacillus cereus</i> ,	It had a strong	[18]

			<i>Micrococcus leutus, Staphylococcus aureus, Shigella Sonni, Aeromonas hydrophilla</i>	antimicrobial effect on most tested bacteria specially on <i>A. hydrophilla</i> then <i>S. sonni</i>	
	1 and 3mg/ml	Myrtacine (Ethanolic extract of leaves)	<i>Erythromycin sensible (EryS) and resistant (EryR) P. acnes strains</i>	Compared with erythromycin, Myrtacine activity was much higher on the EryR strain	[57]
Activity	Dose	Extract/ Constituent	Model	Results	Reference
Antiviral (antiherpetic)	IC ₅₀ before cellular attachment 3.1 mg/ml, and after entering the cells 1.11 mg/ml	Hydroalcoholic extract	The target was Herpes simplex virus-1 (HSV-1) <i>In vitro</i>	By increasing the extract concentration, percentage of inhibition of cytopathic effect was increased	[40]
Anti-diarrhoeal	100, 200 and 400 mg/kg	Methanol extract	Castor oil induced diarrhoea in mice	At dose 200 & 400 mg/kg, the extract significantly delayed the onset of diarrhea	[62]
	5 and 10 ml/kg bw	MBJ	Castor oil-induced	Acute pre-treatment	[63]

	orally		diarrhoea in rat	with MBJ delayed the onset of diarrhea and also decrease the frequency and severity of defecation	
	25 to 100 mg/kg bw orally	Aqueous extract of berries	Castor oil-induced diarrhoea in rat	The extract induced a significant dose-dependent protection against diarrhoea and intestinal fluid accumulation	[64]
Activity	Dose	Extract/ Constituent	Model	Results	Reference
Antidiabetic	500 to 1000 mg/kg bw orally	Aqueous and methanolic extracts	Alloxan-induced diabetic mice	Aqueous extract significantly lowered mean blood glucose level at dose of 500 mg/kg by 61.8%	[65]
	2 g/kg orally	Ethanol-water extract	Streptozotocin-induced diabetes in mice	Significantly reduced the hyperglycaemia	[66]
Antinociceptive	Aqueous extract: 5 to 200	Aqueous and	Hot plate and	Both extracts showed	[67]

	mg/kg i.p., Ethanolic extract: 150 to 350 mg/kg i.p.	Ethanolic Extracts of aerial parts	writhing tests in mice	Significant antinociceptive activity against acetic acid induced writhing and in Hot plate test	
	50 to 150 mg/kg i.p.	Essential oil	Acetic acid induced writhing test in mice	The oil showed dose dependent analgesic effect in comparison with diclofenac sodium	[68]
Anti-plasmodial	1–100 µg/ml	Essential oils	Chloroquine (CQ)-sensitive and resistant strains of <i>P. falciparum</i>	Significant activity present against CQ-sensitive and the resistant strains of <i>P.falciparum</i>	[69]
Activity	Dose	Extract/ Constituent	Model	Results	Reference
Anti-helmintic	0.78 to 50 mg/ml	Ethanolic and water extracts of leaves	<i>In vitro</i> naturally infected cattle using the egg hatch and larval mortality assay	Both extracts have a potential anthelmintic activity on eggs and larvae of bovine strongly parasites	[70]

Oesophageal reflux (ER)		- Aqueous extract of seeds	ER-Induced damage in oesophageal mucosa of adult male Wistar rats	- Aqueous extract exerted a potential protective effect against ER-induced damage in rat oesophagus due to its antioxidant properties	[71]
Antifungal	20 mg/ml	Flavonoids, Terpenoids, of the leaves	<i>Aspergillus</i> species	The highest percentage of inhibition (85%) was recorded at 20 mg/ml of Flavonoids	[11]
	Dose-dependent manner with $\text{MIC}_{50} < 8$ to $64 \mu\text{g}/\text{ml}$.	Oenothein B isolated from seeds	<i>In vitro</i> <i>Candida</i> strains: <i>C. albicans</i> <i>C. glabrata</i> <i>C. parapsilosis</i> <i>C. tropicalis</i>	Oenothein B was able to inhibit the <i>Candida</i> growth.	[72]

Figure 1: *M. communis* plantFigure 2: Flowers of *M. communis*



Figure 4: Seeds of *M. communis*

Figure 3: Leaves of *M. communis*



Figure 5: Berries of *M. communis*



REFERENCES

- [1] Taheri, A., Seyfan, A., Jalalinezhad, S., & Nasery, F. (2013). Antibacterial effect of *Myrtus communis* hydro-alcoholic extract on pathogenic bacteria. *Zahedan J Res Med Sci*, 15(6), 19-24.
- [2] Sumbul, S., Ahmad, M. A., Asif, M., & Akhtar, M. (2011). *Myrtus communis Linn.-A review*.
- [3] Hakeem, M. A., & Mufradat, B. (1895). *Idara Tarraqi* Urdu Publications, Lucknow, p278.
- [4] Fani, M. M., Kohanteb, J. and Araghizadeh, A. (2014). Inhibitory activity of *Myrtus communis* oil on some clinically isolated oral pathogens. *Med Princ Pract*, 23(4), 363-8.
- [5] Alipour, G., Dashti, S. and Hosseinzadeh, H. (2014). Review of pharmacological effects of *Myrtus communis* L. and its active constituents. *Phytotherapy research*, 28(8), 1125-1136.
- [6] Mahmoudvand, H., Ezzatkhah, F., Sharififar, F., Sharifi, I. and Dezaki, E. S. (2015). Antileishmanial and cytotoxic effects of essential oil and methanolic extract of *Myrtus communis* L. *Korean J Parasitol*, 53(1), 21-7.
- [7] Daneshfard, B., Yekta, N. H., Khoshdel, A., Heiran, A., Cheraghi, R. and Yarmohammadi, H. (2019). The effect of *Delphinium denudatum* (Jadwar) on fatigue: A randomized double blind placebo-controlled clinical trial. *Complement Ther Med*, (11), 160-5.
- [8] Jabri, M. A., Marzouki, L. and Sebai, H. (2018). Ethnobotanical, phytochemical and therapeutic effects of *Myrtus communis* L. berries seeds on gastrointestinal tract diseases: a review. *Arch Physiol Biochem*, 124(5), 390-6.
- [9] Abusaief, H. M. A. (2017). A Seed bank study of rare and endangered plants in some locations at Al-Jabal Al-Akhdar of Libya. *Curr. Sci. Int.*, 6(3), 511-539.
- [10] Aleem, M., & Anis, M. (2021). Therapeutic potential of Habb-ul-Aas (*Myrtus communis* Linn.) with Unani Perspective and Modern Pharmacology: A review. *Journal of Pharmacognosy and Phytochemistry*, 10(1), 910-923.

- [11] Hussain, A. Y., Hussein, H. J., & Al-Rubaye, A. F. (2021). Antifungal Efficacy of the crude Flavonoid, Terpenoid, and Alkaloid Extracted from *Myrtus communis* L. against *Aspergillus* species isolated from Stored Medicinal plants seeds in the Iraqi Markets. Journal of Biotechnology Research Center, 15(2), 73-80.
- [12] Eds Satyavati, G. V., Raina, M. K., & Sharma, M. (1976). Medicinal Plants of India Indian council of Medical Research. New Delhi, 4.
- [13] Mitra, R. (1998). Ethno-economic significance of the economic Myrtle-a plant sacred to Greeks and Romans. Ethnobotany, 10(1&2), 1-5.
- [14] Agarwal, V. S. (1986). Economic Plants of India, Kailash Prakashan, Calcutta, p. 251.
- [15] The Wealth of India. (1962). A Dictionary of Indian Raw Materials and Industrial Products ¾ Raw Materials Series, Publications and Information Directorate, Council of Scientific & Industrial Research, New Delhi, India, Vol. VI, pp.482-483.
- [16] Shah, C. S. and Qadri, J. S. (1971). A Textbook of Pharmacognosy, 11th Edn, B.S. Shah Prakashan, Ahmedabad, p. 27.
- [17] Maheshwari, P. and Singh, U. (1965). Dictionary of Economic Plants vof India, Indian Council of Agricultural research, New Delhi, p. 110.
- [18] الناجي عبد الرزاق. & فوزية عبد الرحمن. على سالمة محمود, ادريس (2014). CHEMICAL, PHYSICAL AND MICROBIOLOGICAL STUDIES ON WILD Myrtus communis L. IN EL-JABAL EL-AKDAR AREA-LIBYA الخواص الطبيعية لنبات Myrtus communis L. والكيميائية والميكروبيولوجية لثمار نبات المرسين النامية في منطقة الجبل الأخضر -ليبيا. *Journal of Food and Dairy Sciences*, 5(6), 329-338..
- [19] Haciseferogullari, H., Ozcan, M. M., Arslan, D., & Unver, A. (2012). Biochemical compositional and technological characterizations of black and white myrtle (*Myrtus communis* L.) fruits. *Journal of food science and technology*, 49(1), 82-88.
- [20] Hashemipour, M. A., Lotfi, S., Torabi, M., Sharifi, F., Ansari, M., Ghassemi, A., & Sheikhshoae, S. (2017). Evaluation of the effects of three plant species (*Myrtus Communis* L., *Camellia Sinensis* L., *Zataria Multiflora* Boiss.) on the healing process of intraoral ulcers in rats. *Journal of Dentistry*, 18(2), 127.
- [21] The Useful Plants of India. (1992). Publications and Information Directorate, CSIR, New Delhi, 1992, p. 390.
- [22] Chopra, R. N. (1956). Glossary of Indian medicinal plants.
- [23] Maccioni, S., Tomei, P. E. and Rizzo, A. (1994-1995). L'uso medicinal delle specie vegetali selvatiche coltivate nella tradizione popolare della bassa Val di Magra, Memorie dell'Accademia Lunigianese di Scienze, 64-65, 389-435.
- [24] Yadegarinia, D., Gachkar, L., Rezaei, M. B., Astanch, S. A. and Rasooli, I. (2006). Biochemical activities of Iranian *Mentha piperita* and *Myrtus communis* L. essential oils, *Phytochemistry*, 67, 1249-1255.
- [25] Eds Satyavati , G. V., Raina, M. K., & Sharma, M. (1976). Medicinal Plants of India Indian council of Medical Research. New Delhi, 4.

- [26] Baitar ZI, Aljameul Mufradat Al-advia-wa- al-Aghzia. (1999). Translated by CCRUM, New Delhi, Vol. 1 , pp. 42-47.
- [27] Trease, W. and Evans, D. (2006). Pharmacognosy, 15thEdn, W.B. Saunders Comp Ltd., Toronto, p. 477.
- [28] Mitra, R. (1998). Ethno-economic significance of the economic Myrtle-a plant sacred to Greeks and Romans,Ethnobotany, 10 (1&2), 1-5.
- [29] Kabiruddin M,Makhzan-ul-Mufradat, Sheikh Mohammad Bashir and Sons, Lahore, akistan, (1951). pp. 47-48.
- [30] Serce, S., Ercisli, S., Sengul, M., Gunduz, K., & Orhan, E. (2010). Antioxidant activities and fatty acid composition of wild grown myrtle (*Myrtus communis L.*) fruits. *Pharmacognosy magazine*, 6(21), 9-12.
- [31] Diaz, A. M. and Abeger, A. (1986). Quantitative determination of contents of tannins from the seeds of *Myrtus communis* Linn, Anr Acad Farm, 52(1), 117-122.
- [32] Chandpuri K,Moojizal Qanoon(Urdu translation), Lahore Print Aids, Jama Masjid, Delhi, 1988, pp. 344-345.
- [33] Ghani M N,Khazainul Advia, Sheikh Mohammad Bashir and Sons Publication, Urdu Bazar, Lahore, (1920.) Vol. III, pp. 444-445.
- [34] Chalchat, J., Garry, R. P. and Michet, A. (1998). Essential oils of myrtle of the mediterranean littoral, *J Essent Oil Res*, 10, 613-617.
- [35] Baytop, T. (1999). Therapy with medicinal plants in Turkey (past and present). Publication of the istanbul University, 312.
- [36] Baharvand-Ahmadi, B., Bahmani, M., Naghdi, N., Saki, K., Baharvand-Ahmadi, S., & Rafieian-Kopaei, M. (2015). Review on phytochemistry, therapeutic and pharmacological effects of myrtus (*Myrtus communis*). *Der Pharmacia Lettre*, 7(11), 160-165.
- [37] Al-Traboulsi, M. & Alaib, M. A. (2021).A survey of medicinal plants of Wadi Al-Kouf in Al-Jabal Al-Akhdar, Libya Nat. Croat. Vol. 30(2), 389-404.
- [38] Shaheen, F., Ahmad, M., Nahar Khan, S., Samreen Hussain, S., Anjum, S., Tashkhodjaev, B., ... & Choudhary, M. I. (2006). New α -Glucosidase Inhibitors and Antibacterial Compounds from *Myrtus communis* L.
- [39] Romani, A., Pinelli, P., Mulinacci, N., Vincieri, F. F., & Tattini, M. (1999). Identification and quantitation of polyphenols in leaves of *Myrtus communis* L. *Chromatographia*, 49(1), 17-20.
- [40] Mahmoudvand, H., Ezzatkah, F., Sharififar, F., Sharifi, I., & Dezaki, E. S. (2015). Antileishmanial and cytotoxic effects of essential oil and methanolic extract of *Myrtus communis* L. *The Korean journal of parasitology*, 53(1), 21.
- [41] Jerkovic, I., Radonic, A., & Borcic, I. (2002). Comparative study of leaf, fruit and flower essential oils of Croatian *Myrtus communis* (L.) during a one-year vegetative cycle. *Journal of Essential Oil Research*, 14(4), 266-270
- [42] Nadkarni, K. M. (1989). Indian *Materia Medica*, 3rd Edn, Popular Prakashan Pvt. Ltd., Bombay, vol. 1, p. 838.

- [43] Sastri, B. N. (1962). The Wealth of India. A Dictionary of Indian Raw Materials and Industrial Products. Raw Materials, Vol. 6: LM. *The Wealth of India. A Dictionary of Indian Raw Materials and Industrial Products. Raw Materials, Vol. 6: LM.*
- [44] Montoro, P., Tuberoso, C. I., Perrone, A., Piacente, S., Cabras, P., & Pizza, C. (2006). Characterisation by liquid chromatography-electrospray tandem mass spectrometry of anthocyanins in extracts of *Myrtus communis* L. berries used for the preparation of myrtle liqueur. *Journal of Chromatography A*, 1112(1-2), 232-240.
- [45] Hinou, J., Lakkas, N., & Philianos, S. (1988). Les constituants polyphenoliques de *Myrtus communis* L. *Plant Med Phytoter*, 22, 98-103.
- [46] Martín, T., Rubio, B., Villaescusa, L., Fernández, L., & Díaz, A. M. (1999). Polyphenolic compounds from pericarps of *Myrtus communis*. *Pharmaceutical Biology*, 37(1), 28-31.
- [47] E Derwich , Z Benzianc , R Chabir & R Journal. 3 (3) (2011) 17-23.
- [48] Asif, H. M., Akram, M., Uddin, S., Hasan, Z. U., Sami, A., Iqbal, A., & Tauseef, U. (2011). *Myrtus communis* Linn.(Pharmacological activity). *Journal of Medicinal Plants Research*, 5(26).
- [49] Diaz, A. M., & Abeger, A. (1987). Phenolic compounds of the seeds of *Myrtus communis* L., *Plant Med Phytother*, 21(4), 317-322
- [50] Rastogi, R. P. and Mehrotra, B. N. (1991). Compendium of Indian Medicinal Plants (1970-1979), Central Drug Research Institute Lucknow, Vol. 2, Publications and Information Directorate, CSIR, New Delhi, p. 478.
- [51] Stevens, N. (2005). Natural Synergy: Essential Oils in Cancer Research. http://www.young_living.net/Presentations/Stevens_EssentialOilsInCancer.pdf. Accessed August 1, 2014.
- [52] Bouaziz, A., Abdalla, S., Baghiani, A., & Charef, N. (2015). Phytochemical analysis, hypotensive effect and antioxidant properties of *Myrtus communis* L. growing in Algeria. *Asian Pacific Journal of Tropical Biomedicine*, 5(1), 19-28.
- [53] Hayder, N., Bouhlel, I., Skandrani, I., Kadri, M., Steiman, R., Guiraud, P., ... & Chekir-Ghedira, L. (2008). In vitro antioxidant and antigenotoxic potentials of myricetin-3-o-galactoside and myricetin-3-o-rhamnoside from *Myrtus communis*: Modulation of expression of genes involved in cell defence system using cDNA microarray. *Toxicology in vitro*, 22(3), 567-581.
- [54] Haciseferogullari, H., Özcan, M. M., Arslan, D., & Ünver, A. (2012). Biochemical compositional and technological characterizations of black and white myrtle (*Myrtus communis* L.) fruits. *Journal of food science and technology*, 49(1), 82-88.
- [55] Gardeli, C., Vassiliki, P., Athanasios, M., Kibouris, T., & Komaitis, M. (2008). Essential oil composition of *Pistacia lentiscus* L. and *Myrtus communis* L.: Evaluation of antioxidant capacity of methanolic extracts. *Food chemistry*, 107(3), 1120-1130.
- [56] Janbaz, K. H., Nisa, M., Saqib, F., Imran, I., Zia-Ul-Haq, M., & De Feo, V. (2013). Bronchodilator, vasodilator and spasmolytic activities of methanolic extract of *Myrtus communis* L. *L. J Physiol Pharmacol*, 64(4), 479-84.

- [57] Fiorini-Puybaret, C., Aries, M. F., Fabre, B., Mamatas, S., Luc, J., Degouy, A., ... & Poli, F. (2011). Pharmacological properties of Myrtacine® and its potential value in acne treatment. *Planta medica*, 77(14), 1582-1589.
- [58] Maxia, A., Frau, M. A., Falconieri, D., Karchuli, M. S., & Kasture, S. (2011). Essential oil of *Myrtus communis* inhibits inflammation in rats by reducing serum IL-6 and TNF- α . *Natural product communications*, 6(10), 1545-1548.
- [59] Rossi, A., Di Paola, R., Mazzon, E., Genovese, T., Caminiti, R., Bramanti, P., ... & Cuzzocrea, S. (2009). Myrtucommulone from *Myrtus communis* exhibits potent anti-inflammatory effectiveness in vivo. *Journal of pharmacology and experimental therapeutics*, 329(1), 76-86.
- [60] Alwan, A. H., Al-Gaillany, K. A., & Naji, A. (1989). Inhibition of the binding of 3H-benzo [a] pyrene to rat liver microsomal protein by plant extracts. *International Journal of Crude Drug Research*, 27(1), 33-37.
- [61] Romeilah, R. M. (2016). Chemical compositions, antioxidant, anticancer activities and biological effects of *Myrtus communis* L. and *Origanum vulgare* essential oils. *Asian Journal of Biochemistry*, 11(2), 104-117.
- [62] Sisay, M., Engidawork, E., & Shibeshi, W. (2017). Evaluation of the antidiarrheal activity of the leaf extracts of *Myrtus communis* Linn (Myrtaceae) in mice model. *BMC Complementary and Alternative Medicine*, 17(1), 1-11.
- [63] Jabri, M. A., Rtibi, K., Ben-Said, A., Aouadhi, C., Hosni, K., Sakly, M., & Sebai, H. (2016). Antidiarrhoeal, antimicrobial and antioxidant effects of myrtle berries (*Myrtus communis* L.) seeds extract. *Journal of pharmacy and pharmacology*, 68(2), 264-274.
- [64] Jabri, M. A., Rtibi, K., Sakly, M., Marzouki, L. and Sebai, H. (2016). Role of gastrointestinal motility inhibition and antioxidant properties of myrtle berries (*Myrtus communis* L.) juice in diarrhea treatment. *Biomed Pharmacother*, 84, 1937-44.
- [65] Issa, I. A., & Bule, M. H. (2015). A comparative study of the hypoglycemic effect of aqueous and methanolic extracts of *Myrtus communis* on alloxan induced diabetic Swiss albino mice. *Med Aromat Plants*, 4(190), 2167-0412.
- [66] Elfellah, M. S., Akhter, M. H., & Khan, M. T. (1984). Anti-hyperglycaemic effect of an extract of *Myrtus communis* in streptozotocin-induced diabetes in mice. *Journal of ethnopharmacology*, 11(3), 275-281.
- [67] Hosseinzadeh, H., Khoshdel, M., and Ghorbani, M. (2011). Antinociceptive, Anti-inflammatory Effects and Acute Toxicity of Aqueous and Ethanolic Extracts of *Myrtus communis* L. Aerial Parts in Mice. *JAMS J Acupunct Meridian Stud*, 4(4):242-7.
- [68] Mubarak, S. S., Ibrar, M., Barkatullah, M. N., & Ehsan, M. (2012). Evaluation of essential oil of *Myrtus communis* leaves for analgesic and gastrointestinal motility profile. *Pharmacologyonline*, 2(7), 41-5.
- [69]. Dell'Agli, M., Sanna, C., Rubiolo, P., Basilico, N., Colombo, E., Scaltrito, M. M., ... & Bosisio, E. (2012). Anti-plasmodial and insecticidal activities of the essential

oils of aromatic plants growing in the Mediterranean area. *Malaria journal*, 11(1), 1-10.

[70] Moussouni, L., Besseboua, O. and Ayad, A. (2019). Anthelmintic activity of aqueous and ethanol extracts of *Urtica dioica* L and *Myrtus communis* L Leaves on bovine digestive strongyles: In vitro study. *Ataturk Univ Vet Bilim Derg*, 14(3):273-83.

[71] Jabri, M. A., Tounsi, H., Rtibi, K., Marzouki, L., Sakly, M., & Sebai, H. (2016). Ameliorative and antioxidant effects of myrtle berry seed (*Myrtus communis*) extract during reflux-induced esophagitis in rats. *Pharmaceutical biology*, 54(9), 1575-1585.

[72] Franco, A. M., Tocci, N., Guella, G., Dell'Agli, M., Sangiovanni, E., Perenzoni, D., ... & Manca, G. (2019). Myrtle Seeds (*Myrtus communis* L.) as a rich source of the bioactive ellagitannins oenothein B and eugeniflorin D2. *ACS omega*, 4(14), 15966-15974.